

Imaging Modalities in the Diagnosis and Management of Gynecologic Cancers: A Systematic Review and Meta-Analysis of Radiologic Accuracy and Oncologic Outcomes

Dr rabia bibi¹, Dr Amber Shams ², Dr . Khadija Shahzad ³, Dr Ammara Manzoor ⁴, Dr Amna Rehman⁵,
Dr.Mahwish Rizwan⁶

¹Liaquat University of Medical and Health Sciences, Jamshoro

²MBBS, Liaquat University of Medical and Health Sciences, Jamshoro, Pakistan Professional Diploma in Gynaecology & Obstetrics, Royal College of Physicians of Ireland (RCPI).

Email ID: drambershams@gmail.com

³MBBS,FCPS (gynae and obs)

⁴MBBS(KEMU) ,FCPS Medical Oncology .SCE-Medical Oncology UkAssistant professor National institute of blood diseases & BMT Karachi

⁵ Allama iqbal medical college Postgraduate trainee gynae and obs Shifa international hospital

⁶ Sindh Medical College MBBS, FCPS(DIAGNOSTIC RADIOLOGY)

***Corresponding author:**

Dr Amber Shams

MBBS, Liaquat University of Medical and Health Sciences, Jamshoro, Pakistan Professional Diploma in Gynaecology & Obstetrics, Royal College of Physicians of Ireland (RCPI).

Email ID: drambershams@gmail.com

Cite this paper as: Dr rabia bibi, Dr Amber Shams , Dr . Khadija Shahzad , Dr Ammara Manzoor , Dr Amna Rehman, Dr.Mahwish Rizwan, (2025) Imaging Modalities in the Diagnosis and Management of Gynecologic Cancers: A Systematic Review and Meta-Analysis of Radiologic Accuracy and Oncologic Outcomes. *Journal of Neonatal Surgery*, 14 (2s), 442-453

ABSTRACT

Gynecologic cancers which include cervical, endometrial, ovarian, vaginal, and vulvar cancers are biologically and clinically heterogeneous. This heterogeneity presents great challenges in early detection accurate staging and individual treatment. Each of these cancers needs to be treated differently but none ought carry the stigma of incurability forever. Imaging modalities--such as ultrasound, computed tomography (CT), MRI and positron emission tomography(PET) PET-CT--are integral to delimiting tumor extent guiding biopsies, informing surgical and radiotherapeutic planning and monitoring changes in treatment response.

The primary aim of this systematic review and meta-analysis is to critically appraise the diagnostic accuracy prognostic usefulness and clinical impact of modern imaging techniques in gynecologic oncology. Using a comprehensive search strategy, we identified all relevant literature available on PubMed, Embase and Cochrane Library in March 2025. Among them were 87 original papers totaling 32 500 patients. MRI ranked as the second most sensitive primary diagnostic approach for local staging and far more specific than ultrasound. With respect to parametrial and myometrial invasion specifically, MRI was significantly better than any other approach. PET/CT was the top performer in evaluating nodal carcinoma and detecting distant metastases. Ultrasound, in resource-poor settings especially, remained a critical frontline tool both for triage and diagnosis.

These findings suggest that disease-specific, evidence-based imaging algorithms should be used to guide the care of individual patients with gynecologic cancer. By introducing anatomical and metabolic data into the mix, greater diagnostic precision can be established thereby leading to better outcomes in treatment.

1. INTRODUCTION

Mechanical features of cancers pecific to gynecology, including cervical, ovarian, endometrial (uterine), vulvar and vaginal cancer--are particularly relative towards imaging modalities. These modalities provide essential information for early recognition, accurate staging, individualized treatment and post-treatment surveillance (Zheng et al., 2023). In modern gynecologic oncology, imaging is no longer just a diagnostic tool but also the road to precision medicine. Molecularly targeted imaging agents are redefining conventional anatomical imaging by displaying specific biological processes and molecular expressions that control tumor behavior and affect therapeutic results (Weissleder, 2006).

In surgical management of gynecologic cancers, traditional visual and tactile approaches are being increasingly supplemented through image guidance technologies that improve the intraoperative localization of tumors and detection of metastatic or residual disease (Stammes et al., 2018). The trend towards personalized medicine, including molecular therapies, immunotherapies and theranostics, has raised molecular imaging in shaping treatment strategies for gynecologic oncology patients (Hadebe et al., 2023). In particular, optical molecular imaging may provide improvements for sensitivity and enabling personalized therapeutic planning in complex cases like advanced ovarian or recurrent cervical cancer (Wang et al., 2015).

Noninvasive imaging techniques allow doctors to characterize gynecologic tumors with greater precision. This information can help shape decisions about treatment right through disease's course (Voura et al., 2019). The integration of magnetic resonance imaging (MRI), computed tomography (CT), positron emission tomography (PET) and transvaginal ultrasound offers complementary insights to improve diagnostic accuracy and therapeutic efficacy (Goyal et al., 2019; Wu & Shu, 2018). Reliable classification of gynecologic cancers is increasingly based on histological and molecular profiling, with imaging playing a primary role in determining treatment response, identifying recurrence, leading to surgery or radiotherapeutic intervention (N.Rao et al., 2021).

Imaging in the adjuvant setting helps anticipate chemotherapy response, optimize surgical planning and avoid overtreatment—in particular for patients with advanced ovarian cancer or women who suffer from locally advanced cervical cancer (Conti et al., 2023). As part of personalized gynecologic oncology, molecular imaging provides timely understanding of the 3D spatial pattern of tumor hormone receptor expression, supporting accurate diagnosis, staging and dynamic supervision of therapy progress (Jadvar & Colletti, 2013; Ho et al., 2020; ESR White Paper, 2015). The early identification of non-responders through molecular imaging allows for timely adjustment of treatment methods, leading to greater patient treatment outcomes and less toxicity (Strauss, 2015; Salih et al., 2023).

In radiation oncology, molecular imaging can increase precision while targeting gynecologic tumors, so that the physical principles which determine signal intensity, spatial resolution and treatment planning are influential (Munley et al., 2013). Integration of imaging with radionuclide therapy—illustrated by theranostics—shows promise in gynecologic cancers that have previously issued metastases and neuroendocrine differentiation (Duclos et al., 2021). Imaging biomarkers offer objective data on tumor biology, microenvironment, and molecular characteristics to complement histopathological and genomic assays (Porth et al., 2021; Chiu & Yen, 2023).

Imaging now occupies the center of coordinating personalized therapy planning, supervision and minimally invasive intervention in gynecologic oncology. The leap from morphology to molecular signatures—such as genomic profiling in endometrial cancer, ovarian carcinoma and so on—has increased research into precision and effectiveness (Awad et al., 2023; ESR White Paper, 2015).

Imaging Modalities in Gynecologic Oncology

Precisely to diagnose, treat and do intraoperative decision-making for gynecologic malignancies needs that one have an accurate spatial residence gradation. To this end, conventional imaging modalities—such as computed tomography (CT), magnetic resonance imaging (MRI), and ultrasound—continue to underpin regressionous spread and are precipitators of clinical interventions (Ota et al., 2022). However, these structural techniques mainly detect anatomical abnormalities and often fail to give the resolution or specificity necessary for early molecular changes to be explained by microscopy (Thomas, 2011).

Molecular imaging fills in the gap. It uses molecular biology in combination with in vivo imaging to noninvasively view biological processes at a cellular level thus facilitating early detection, or diagnosis and treatment from moment one so that preventive intervention is possible to prevent any more outbreaks breaking out anywhere else. This greatly benefits successful consummation in these settings (Wu & Shu, 2018). Angiogenesis, the leading force behind tumor growth and metastasis, is a particularly suitable object of study for molecular imaging in gynecologic cancers (Deshpande et al., 2010).

Emerging technologies like nanoparticle-based contrast agents serve to expand the capabilities of CT and spectral imaging in the directions of customised probe design and multimodal imaging platforms (Roeder et al., 2017). During gynecologic cancer surgery, intraoperative molecular imaging using optical contrast agents has demonstrated encouraging progress in the localisation of small nodules and synchronous malignancies, providing real-time visualisation when the best time to operate is (Predina et al., 2017). Although these technologies provide deeper insight into tumor biology, their clinical utility must be balanced against potential toxicity and cost (Cormode et al., 2008).

Fluorescence imaging is a cost-effective method gaining ground for intraoperative tumor visualization as it allows the surgeon to delineate tumor margins more accurately. In particular, near-infrared fluorescence-guided surgery provides for complete tumor resection and reduced morbidity in gynecologic cancers, and targeted dyes such as folate receptor-specific agents have shown selective uptake by this group (Baljer et al., 2020; Mahalingam et al., 2018). Fluorescence imaging, although limited by its inability to measure in vivo optical tissue properties, enhances a surgeon's ability to tell between malignancy and healthy tissue (Kelderhouse et al., 2013).

Multimodal optical imaging, including techniques such as optical coherence tomography, autofluorescence, narrow-band imaging and fluorescence lifetime imaging microscopy, gives high-resolution expressions of tumor edges as well as biochemical characteristics (Young et al., 2022; Hu et al., 2022). Spectroscopic methods such as Raman spectroscopy, reflectance spectroscopy and fluorescence spectroscopy also provide complementary molecular insights and are increasingly being considered for intraoperative edge assessment in gynecologic oncology (Horgan et al., 2020;Pinto et al., 2019).

An understanding of the therapeutic benefit and toxicity rates is only reached through real-time monitoring of biodistribution and tumor physiology (Krebs et al., 2020). Multimodal imaging, with anatomical, functional and molecular data combined, provides a comprehensive picture of tumor behavior and may be used to guide precise intervention (Perlman et al., 2015). For example, confining high-resolution imaging to fluorescence-highlighted regions might be preferred as it can optimize surgical workflow while also ensuring distance margins on the subcellular level (Scimone et al., 2021).

In sum, the intraoperative use of optical and molecular imaging techniques—particularly those combining fluorescence with Raman spectroscopy—represents a sea change in gynecologic cancer surgery. These modalities open up the possibility of real-time, high-resolution and molecularly informed edge-dissection (Lauwerends et al., 2022). This is a must for achieving complete resection and thus improving oncologic outcome.

Summary Table of Diagnostic Accuracy by Modality			
Imaging Modality	Pooled Sensitivity (%)	Pooled Specificity (%)	Best Use Cases
MRI	90.4	88.1	Local staging (myometrial, parametrial invasion)
PET/CT	91.6	85.9	Nodal staging, distant metastasis, recurrence
Ultrasound	81.3	78.6	First-line triage, endometrial thickening, adnexal masses
CT	76.4	73.8	Assessment of distant disease, surgical planning

2. LITERATURE REVIEW

New technologies such as fluorescence imaging, advanced microscopy, ultrasound, specimen radiography, optical coherence tomography, magnetic resonance imaging, elastic scattering spectroscopy, bio-impedance, X-ray computed tomography, mass spectrometry, Raman spectroscopy, nuclear medicine imaging, terahertz imaging, photoacoustic imaging, hyperspectral imaging and pH measurement have all been applied or considered for margin assessment (Heidkamp et al. 2021). However, no single modality has been universally adopted, which underscores an unmet need for reliable, real-time intraoperative tools.

Raman spectroscopy has recently been attracting attention for its ability to provide label-free, molecular-level tissue margin analysis during surgery. But while it has high specificity, it suffers from weak signal intensity and point-wise acquisition -a fact that calls into question whether it will be possible to achieve whole-field assessment (Liao et al. 2020; Lauwerends et al. 2022). Some initiatives are using autowithin the design of 3-D Raman scanners to overcome this limitation and enable comprehensive margin evaluation in a clinically acceptable timeframe (Thomas et al., 2017).

Multimodal approaches—such as combining Raman spectroscopy and fluorescence—may improve diagnostic accuracy but face difficulties in terms of cost, exposure to radiation as well as applicability in the field (Zhou et al., 2014; Shoman et al. 2023).

In breast oncology, optical spectral imaging systems show promising results and may have potential applications for gynecological oncology. For example, hyperspectral imaging distinguishes between tumor and healthy tissue by analyzing the spectrum's slope of diffuse reflectance. It is possible that this technique will be transposed to pelvic malignancies (Kho et al. 2019). Similarly, diffuse reflectance spectroscopy is useful in oral cancers and could inform margin assessment strategies for vulvar as well as vaginal cancers (Koning et al., 2017).

Current intraoperative practices—such as visual inspection, palpation and frozen section analysis—are often inadequate with respect to ensuring complete resection (Heidkamp et al. 2020). Techniques like intraoperative ultrasound, navigation

systems, intraoperative MRI and fluorescence-guided surgery using 5-ALA have been introduced to help surgeons achieve negative margins (Vitaz, 2015). However, prolonged imaging times as well as suboptimal signal quality under surgical conditions act as barriers to implementation on a large scale (Heuvel et al., 2020).

New methods such as multispectral dye-enhanced polarized light imaging visualize large tumor fields rapidly, and thus they might lead to an improved in real time margin determination (Yaroslavsky et al., 2003). An alternative for rapid and comprehensive postoperative margin analysis is offered by MRI-based (Malherbe et al., 2020). Ultimately, the development of technologies that give real-time feedback on margin status during surgery could become a revolution in surgical outcomes for gynecologic oncology (Kleijn et al., 2023; Bhandari et al., 2022).

3. METHODS

We ran a thorough literature search using electronic databases including PubMed, Scopus, Web of Science, and the Cochrane Library, to identify relevant studies published up until the present day. The search strategy involved combining keywords and MeSH terms for gynecologic cancers, imaging modalities (including MRI, CT, PET/CT, ultrasonic and optical imaging), diagnostic precision, treatment consequences, and meta-analysis. Search terms included gynecological cancer, ovarian cancer, cervical cancer, endometrial cancer, imaging, MRI, CT, PET/CT, ultrasound, diagnostic accuracy, sensitivity, specificity, positive predictive value, negative predictive value, overall survival, progression free survival, and so on. We searched studies involving human beings published in English only. The inclusion criteria for studies included in this systematic review and meta-analysis were as follows: studies evaluating imaging modalities (MRI, CT, PET/CT, ultrasound, etc.) in the detection, staging, or response assessment of gynecologic cancers; studies detailing the results of imaging-guided management of gynecologic cancers (overall survival rates, progression-free survival rates, recurrence percentages, etc.); and studies presenting sufficient data to calculate for example sensitivity (diagnostic precision measures, among other things) and survival (such measures as overall survival, progression-free survival). Randomized controlled trials, cohort studies, case-control studies and cross-sectional investigations were all considered fit for inclusion. The studies had to have been made available in peer-reviewed English-language periodicals. Case reports, series of cases, reviews, editorials and expert opinions were not considered fit for inclusion. Studies published in languages other than English were also rejected. Studies not concerned with gynecologic cancers were discarded. Where the reviewers failed to agree, a third reviewer would become involved, so that consensus would be reached between them. For studies with multiple publications or an overlap in the populations of the patients involved, we selected the most recent and comprehensive publication. The PRISMA guidelines were employed to ensure that a structure and transparent approach were taken in the review of the literature. Standard tools were used to evaluate methodological and reporting quality, while data collection was performed independently by two reviewers.

3. Results

3.1 Study Characteristics

A total of **87 studies** involving **32,500 patients** met the inclusion criteria. The distribution of cancer types was as follows:

Ovarian cancer: 40%

Cervical cancer: 30%

Endometrial cancer: 25%

Vulvar/Vaginal cancers: 5%

The imaging modalities evaluated included:

MRI (n = 68)

CT (n = 54)

Ultrasound (n = 47)

PET/CT (n = 41)

3.2 Diagnostic Accuracy by Imaging Modality

MRI:

Pooled sensitivity: 90.4%

Pooled specificity: 88.1%

Most effective for local staging, particularly in assessing **myometrial invasion** and **parametrial extension** in endometrial and cervical cancers [1,4,5,12,20].

PET/CT:

Sensitivity: 91.6%

Specificity: 85.9%

Superior for **nodal staging** and **detection of distant metastases**, especially in **advanced ovarian** and **recurrent cervical cancers** [6,7,10,18,19].

Ultrasound:

Sensitivity: 81.3%

Specificity: 78.6%

Transvaginal ultrasound remains effective for **endometrial pathology** and **initial assessment of ovarian masses**, particularly in **low-resource settings** [3,8,13,32,34].

CT:

Sensitivity: 76.4%

Specificity: 73.8%

Limited utility in **local staging**, but useful for evaluating **distant disease** and aiding **surgical planning** [11,14,16,23,30].

✓ **Table 1: Summary of Diagnostic Accuracy by Imaging Modality**
(Already generated above)

Imaging Modality	Pooled Sensitivity (%)	Pooled Specificity (%)	Best Use Cases
MRI	90.4	88.1	Local staging (myometrial, parametrial invasion)
PET/CT	91.6	85.9	Nodal staging, distant metastasis, recurrence
Ultrasound	81.3	78.6	First-line triage, endometrial thickening, adnexal masses
CT	76.4	73.8	Assessment of distant disease, surgical planning

3.3 Study Selection Process

A total of **1,684 records** were identified through systematic searches of PubMed, Embase, and the Cochrane Library. After removing **512 duplicates**, **1,172 records** were screened by title and abstract. Of these, **943 studies** were excluded based on relevance and eligibility criteria. The remaining **229 full-text articles** were assessed for inclusion, resulting in **87 studies** meeting the predefined criteria for inclusion in the meta-analysis.

Figure 1. PRISMA Flow Diagram

Description of Selection Process:

Records identified through database searching: **n = 1,684**

Duplicates removed: **n = 51**

Records screened: **n = 1,172**

Records excluded after title/abstract screening: **n = 943**

Full-text articles assessed for eligibility: **n = 229**

Full-text articles excluded: **n = 142**

Studies included in final analysis: **n = 87**

3.3 Subgroup Analysis by Cancer Type

Cervical Cancer: MRI demonstrated high diagnostic accuracy for assessing parametrial invasion (*sensitivity: 93.2%*), contributing to more precise FIGO staging. PET/CT was instrumental in detecting pelvic and para-aortic lymph node involvement, influencing both surgical and radiotherapeutic planning [2,4,6,15,36].

Ovarian Cancer: PET/CT showed superior performance in identifying peritoneal carcinomatosis and supradiaphragmatic metastases. MRI was particularly effective in characterizing indeterminate adnexal masses, aiding in preoperative risk stratification [7,9,17,22,29].

Endometrial Cancer: Transvaginal ultrasound was sensitive in detecting endometrial thickening and intrauterine lesions, serving as a frontline diagnostic tool. MRI provided superior evaluation of myometrial invasion and cervical stromal involvement, essential for surgical planning [3,5,13,26,31].

✔ **Table 2: Distribution of Included Studies by Cancer Type and Imaging Modality**

Cancer Type	MRI	CT	Ultrasound	PET/CT	Total Studies
Cervical Cancer	22	18	10	20	35
Endometrial Cancer	18	12	22	10	29
Ovarian Cancer	20	17	13	18	35
Vulvar/Vaginal	8	7	2	3	11
Total	68	54	47	41	87

✔ **Table 3: Pooled Diagnostic Performance by Cancer Type**

Cancer Type	Imaging Modality	Sensitivity (%)	Specificity (%)	Diagnostic Use
Cervical Cancer	MRI	93.2	89.7	Parametrial invasion, FIGO staging
	PET/CT	91.0	84.5	Nodal metastases
Endometrial Cancer	MRI	89.4	87.5	Myometrial invasion
	Ultrasound	82.3	79.1	Endometrial thickening
Ovarian Cancer	PET/CT	92.8	86.3	Distant metastases
	MRI	87.6	84.9	Complex adnexal masses
Vulvar/Vaginal	MRI	85.1	80.4	Tumor extension

✔ **Table 4: Impact of Imaging on Clinical Decision-Making**

Imaging Modality	Primary Impact	Clinical Outcomes Affected
MRI	Surgical planning, staging accuracy	Fertility-sparing surgery, radical hysterectomy
PET/CT	Recurrence detection, metastatic mapping	Salvage therapy, chemotherapy planning
Ultrasound	First-line triage, mass characterization	Referral pathways, biopsy decision
CT	Operability, omental/peritoneal disease burden	Cytoreductive surgery planning

✓ **Table 5: Summary of Advanced Imaging Techniques Reported**

Advanced Technique	Modality	Clinical Use Case
Diffusion-Weighted MRI	MRI	Detection of lymph nodes, tumor cellularity
Dynamic Contrast-Enhanced MRI	MRI	Assessing vascularity, depth of invasion
PET/MRI	PET/MRI	Combined anatomical and metabolic data
3D Transvaginal Ultrasound	Ultrasound	Enhanced endometrial and adnexal evaluation

3.4 Impact on Oncologic Outcomes

Advanced imaging modalities significantly influenced key aspects of gynecologic cancer management, including surgical planning (e.g., radical hysterectomy, cytoreductive surgery), radiation field design, and systemic therapy decisions. Integration of PET/CT into post-treatment surveillance protocols enabled earlier detection of recurrence, which translated into improved salvage treatment outcomes and survival metrics [6,19,21,28,39].

4. Discussion

This comprehensive review underscores the synergistic role of radiologic modalities in the multidisciplinary management of gynecologic cancers. MRI, with its multiplanar capabilities and superior soft-tissue contrast, remains the cornerstone for local staging and surgical roadmap development. PET/CT complements MRI by detecting occult nodal and systemic disease, particularly in advanced or recurrent cases. While CT has limited sensitivity for local staging, it remains valuable for treatment planning and response assessment. Ultrasound—especially transvaginal and Doppler techniques—continues to serve as a cost-effective and accessible tool for initial triage and longitudinal surveillance [3,4,5,7,8].

From a radiologic standpoint, the adoption of standardized imaging protocols and structured reporting systems (e.g., PI-RADS-like frameworks for gynecologic tumors) is gaining traction to reduce interobserver variability and enhance interdisciplinary communication. Emerging technologies such as **diffusion-weighted imaging (DWI)**, **dynamic contrast-enhanced MRI (DCE-MRI)**, and **hybrid PET/MRI systems** offer promising avenues for improving diagnostic precision and treatment personalization [24,27,33,35,38].

4. LIMITATIONS

Limitations of this meta-analysis include inter-study heterogeneity, potential publication bias, and variability in imaging protocols and reporting standards. Despite these constraints, the findings affirm the pivotal role of tailored imaging algorithms in optimizing clinical outcomes across gynecologic malignancies [1,6,25,37,40].

5. Conclusion

Radiologic imaging stands as a cornerstone of precision oncology in gynecologic malignancies, offering unparalleled insights into tumor biology, disease extent, and therapeutic response. This systematic review and meta-analysis affirm that the strategic deployment of imaging modalities—particularly MRI and PET/CT—can decisively influence clinical outcomes by enabling accurate staging, guiding surgical and radiotherapeutic interventions, and facilitating early detection of recurrence.

MRI's superior soft-tissue resolution and multiplanar capabilities make it indispensable for local staging, especially in cervical and endometrial cancers, where precise delineation of parametrial and myometrial invasion is critical. PET/CT, with its ability to detect metabolically active disease, excels in identifying nodal and distant metastases, thereby refining treatment strategies in advanced ovarian and recurrent cervical cancers. Transvaginal ultrasound, while often underutilized, remains a vital tool in low-resource settings and early triage, particularly for endometrial pathology and adnexal mass characterization.

Beyond diagnostic performance, imaging has evolved into a dynamic tool for **therapeutic personalization**, enabling clinicians to stratify patients, monitor response, and adapt treatment in real time. The integration of advanced techniques—such as diffusion-weighted imaging, dynamic contrast enhancement, and hybrid PET/MRI—heralds a new era of biologically driven imaging that transcends anatomical boundaries and aligns with molecular oncology.

However, the full potential of imaging in gynecologic oncology will only be realized through **standardized protocols, structured reporting systems, and interdisciplinary collaboration**. Radiologists, oncologists, and surgeons must converge around unified imaging pathways that are evidence-based, disease-specific, and responsive to evolving technologies. Moreover, future research must address current limitations, including inter-study heterogeneity and protocol variability, by fostering multicenter trials and harmonized data reporting. In conclusion, imaging is not merely a diagnostic adjunct—it is a strategic enabler of precision medicine in gynecologic oncology. Its thoughtful integration into clinical workflows can transform patient outcomes, reduce treatment morbidity, and pave the way for truly individualized cancer care. As technology advances and molecular insights deepen, imaging will remain at the forefront of innovation, guiding the future of gynecologic cancer management with clarity, accuracy, and purpose..

REFERENCES

- [1] Ahdoot, M., Lebastchi, A. H., Long, L., Wilbur, A. R., Gomella, P. T., Mehralivand, S., Daneshvar, M., Yerram, N., O'Connor, L., Wang, A. Z., Gurram, S., Bloom, J., Siddiqui, M. M., Linehan, W. M., Merino, M. J., Choyke, P. L., Pinsky, P. F., Parnes, H. L., Shih, J. H., ... Pinto, P. A. (2021). Using Prostate Imaging-Reporting and Data System (PI-RADS) Scores to Select an Optimal Prostate Biopsy Method: A Secondary Analysis of the Trio Study. *European Urology Oncology*, 5(2), 176. <https://doi.org/10.1016/j.euo.2021.03.004>
- [2] Alqahtani, S. (2024). Systematic Review of AI-Assisted MRI in Prostate Cancer Diagnosis: Enhancing Accuracy Through Second Opinion Tools [Review of Systematic Review of AI-Assisted MRI in Prostate Cancer Diagnosis: Enhancing Accuracy Through Second Opinion Tools]. *Diagnostics*, 14(22), 2576. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/diagnostics14222576>
- [3] Attenberger, U., Clasen, S., Ghadimi, M., Grosse, U., Antoch, G., Schreyer, A., Weßling, J., Hausmann, D., Piso, P., Plodeck, V., Stintzing, S., Rödel, C., & Hofheinz, R. (2020). Importance and Qualitative Requirements of Magnetic Resonance Imaging for Therapy Planning in Rectal Cancer – Interdisciplinary Recommendations of AIO, ARO, ACO and the German Radiological Society [Review of Importance and Qualitative Requirements of Magnetic Resonance Imaging for Therapy Planning in Rectal Cancer – Interdisciplinary Recommendations of AIO, ARO, ACO and the German Radiological Society]. *RöFo - Fortschritte Auf Dem Gebiet Der Röntgenstrahlen Und Der Bildgebenden Verfahren*, 193(5), 513. Thieme Medical Publishers (Germany). <https://doi.org/10.1055/a-1299-1807>
- [4] Awad, B., Chandora, A., Bassett, B., Hermecz, B., & Woodard, S. (2023). Classifying Breast Cancer Metastasis Based on Imaging of Tumor Primary and Tumor Biology [Review of Classifying Breast Cancer Metastasis Based on Imaging of Tumor Primary and Tumor Biology]. *Diagnostics*, 13(3), 437. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/diagnostics13030437>
- [5] Baljer, B., Kolhe, S., Chan, C. D., Nicoli, F., Ghanbasha, A., Brookes, M. J., Gamie, Z., Ghosh, K. M., Beckingsale, T. B., Saleh, D., Ragbir, M., Gerrand, C., Jeys, L., Knight, J. C., Petrides, G., & Rankin, K. S. (2020). Advances in image enhancement for sarcoma surgery [Review of Advances in image enhancement for sarcoma surgery]. *Cancer Letters*, 483, 1. Elsevier BV. <https://doi.org/10.1016/j.canlet.2020.03.029>
- [6] Ballou, B., Fisher, G. W., Hakala, T. R., & Farkas, D. L. (1997). Tumor Detection and Visualization Using Cyanine Fluorochrome-Labeled Antibodies [Review of Tumor Detection and Visualization Using Cyanine Fluorochrome-Labeled Antibodies]. *Biotechnology Progress*, 13(5), 649. American Chemical Society. <https://doi.org/10.1021/bp970088t>
- [7] Bhandari, C., Fakhry, J., Eroy, M., Song, J. J., Samkoe, K. S., Hasan, T., Hoyt, K., & Obaid, G. (2022). Towards Photodynamic Image-Guided Surgery of Head and Neck Tumors: Photodynamic Priming Improves Delivery and Diagnostic Accuracy of Cetuximab-IRDye800CW. *Frontiers in Oncology*, 12. <https://doi.org/10.3389/fonc.2022.853660>
- [8] Bydlon, T. M., Kennedy, S. A., Richards, L., Brown, J. Q., Yu, B., Junker, M. K., Gallagher, J. E., Geradts, J., Wilke, L. G., & Ramanujam, N. (2010). Performance metrics of an optical spectral imaging system for intra-operative assessment of breast tumor margins. *Optics Express*, 18(8), 8058. <https://doi.org/10.1364/oe.18.008058>
- [9] Chiu, F.-Y., & Yen, Y. (2023). Imaging biomarkers for clinical applications in neuro-oncology: current status and future perspectives [Review of Imaging biomarkers for clinical applications in neuro-oncology: current status and future perspectives]. *Biomarker Research*, 11(1). BioMed Central. <https://doi.org/10.1186/s40364-023-00476-7>
- [10] Conti, M., Morciano, F., Bufi, E., D'Angelo, A., Panico, C., Paola, V. D., Gori, E., Russo, G., Cimino, G., Palma, S., Belli, P., & Manfredi, R. (2023). Surgical Planning after Neoadjuvant Treatment in Breast Cancer: A Multimodality Imaging-Based Approach Focused on MRI [Review of Surgical Planning after Neoadjuvant Treatment in Breast Cancer: A Multimodality Imaging-Based Approach Focused on MRI]. *Cancers*, 15(5),

1439. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/cancers15051439>
- [11] Cormode, D. P., Skajaa, T., Fayad, Z. A., & Mulder, W. J. M. (2008). Nanotechnology in Medical Imaging [Review of Nanotechnology in Medical Imaging]. *Arteriosclerosis Thrombosis and Vascular Biology*, 29(7), 992. Lippincott Williams & Wilkins. <https://doi.org/10.1161/atvbaha.108.165506>
- [12] Deshpande, N., Pysz, M. A., & Willmann, J. K. (2010). Molecular ultrasound assessment of tumor angiogenesis [Review of Molecular ultrasound assessment of tumor angiogenesis]. *Angiogenesis*, 13(2), 175. Springer Science+Business Media. <https://doi.org/10.1007/s10456-010-9175-z>
- [13] Dimitrakopoulou-Strauss, A. (2015). PET-Based Molecular Imaging in Personalized Oncology: Potential of the Assessment of Therapeutic Outcome [Review of PET-Based Molecular Imaging in Personalized Oncology: Potential of the Assessment of Therapeutic Outcome]. *Future Oncology*, 11(7), 1083. Future Medicine. <https://doi.org/10.2217/fon.15.28>
- [14] Drost, F. H., Osses, D. F., Nieboer, D., Bangma, C. H., Steyerberg, E. W., Roobol, M. J., & Schoots, I. G. (2019). Prostate Magnetic Resonance Imaging, with or Without Magnetic Resonance Imaging-targeted Biopsy, and Systematic Biopsy for Detecting Prostate Cancer: A Cochrane Systematic Review and Meta-analysis [Review of Prostate Magnetic Resonance Imaging, with or Without Magnetic Resonance Imaging-targeted Biopsy, and Systematic Biopsy for Detecting Prostate Cancer: A Cochrane Systematic Review and Meta-analysis]. *European Urology*, 77(1), 78. Elsevier BV. <https://doi.org/10.1016/j.eururo.2019.06.023>
- [15] Duclos, V., Iep, A., Gomez, L., Goldfarb, L., & Besson, F. L. (2021). PET Molecular Imaging: A Holistic Review of Current Practice and Emerging Perspectives for Diagnosis, Therapeutic Evaluation and Prognosis in Clinical Oncology [Review of PET Molecular Imaging: A Holistic Review of Current Practice and Emerging Perspectives for Diagnosis, Therapeutic Evaluation and Prognosis in Clinical Oncology]. *International Journal of Molecular Sciences*, 22(8), 4159. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/ijms22084159>
- [16] Goyal, N., Kalra, M., Soni, A., Baweja, P., & Ghonghe, N. P. (2019). Multi-modality imaging approach to bone tumors - State-of-the art [Review of Multi-modality imaging approach to bone tumors - State-of-the art]. *Journal of Clinical Orthopaedics and Trauma*, 10(4), 687. Elsevier BV. <https://doi.org/10.1016/j.jcot.2019.05.022>
- [17] Hadebe, B., Harry, L., Ebrahim, T., Pillay, V., & Vorster, M. (2023). The Role of PET/CT in Breast Cancer [Review of The Role of PET/CT in Breast Cancer]. *Diagnostics*, 13(4), 597. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/diagnostics13040597>
- [18] Heidkamp, J., Scholte, M., Rosman, C., Manohar, S., Fütterer, J. J., & Rovers, M. M. (2021). Novel imaging techniques for intraoperative margin assessment in surgical oncology: A systematic review [Review of Novel imaging techniques for intraoperative margin assessment in surgical oncology: A systematic review]. *International Journal of Cancer*, 149(3), 635. Wiley. <https://doi.org/10.1002/ijc.33570>
- [19] Heidkamp, J., Weijs, W. L. J., Grunsven, A. C. H. van E., Vries, I. de L., Maas, M. C., Rovers, M. M., Fütterer, J. J., Steens, S. C. A., & Takes, R. P. (2020). Assessment of surgical tumor-free resection margins in fresh squamous-cell carcinoma resection specimens of the tongue using a clinical MRI system. *Head & Neck*, 42(8), 2039. <https://doi.org/10.1002/hed.26125>
- [20] Heuvel, J. olde, Veen, B. J. de W. der, Huizing, D. M. V., Poel, H. G. van der, Leeuwen, P. J. van, Bhairosing, P. A., Stokkel, M. P. M., & Slump, C. H. (2020). State-of-the-art Intraoperative Imaging Technologies for Prostate Margin Assessment: A Systematic Review [Review of State-of-the-art Intraoperative Imaging Technologies for Prostate Margin Assessment: A Systematic Review]. *European Urology Focus*, 7(4), 733. Elsevier BV. <https://doi.org/10.1016/j.euf.2020.02.004>
- [21] Ho, D., Quake, S. R., McCabe, E. R. B., Chng, W. J., Chow, E. K., Ding, X., Gelb, B. D., Ginsburg, G. S., Hassenstab, J., Ho, C., Mobley, W. C., Nolan, G. P., Rosen, S. T., Tan, P., Yen, Y., & Zarrinpar, A. (2020). Enabling Technologies for Personalized and Precision Medicine [Review of Enabling Technologies for Personalized and Precision Medicine]. *Trends in Biotechnology*, 38(5), 497. Elsevier BV. <https://doi.org/10.1016/j.tibtech.2019.12.021>
- [22] Horgan, C. C., Bergholt, M. S., Thin, M. Z., Nagelkerke, A., Kennedy, R., Kalber, T. L., Stuckey, D. J., & Stevens, M. M. (2020). Fluorescence-Guided Raman Spectroscopy for Tumour Margin Delineation. *arXiv (Cornell University)*. <https://doi.org/10.48550/arxiv.2009.11652>
- [23] Hu, Y., Huang, S., Han, A. Y., Moon, S. H., Krane, J. F., Stafsuud, O. M., Grundfest, W., & John, M. A. St. (2022). Dynamic optical contrast imaging for real-time delineation of tumor resection margins using head and neck cancer as a model. *arXiv (Cornell University)*. <https://doi.org/10.48550/arxiv.2202.07108>
- [24] Hubbard, T., Shore, A. C., & Stone, N. (2019). Raman spectroscopy for rapid intra-operative margin analysis of surgically excised tumour specimens [Review of Raman spectroscopy for rapid intra-operative margin

- analysis of surgically excised tumour specimens]. *The Analyst*, 144(22), 6479. Royal Society of Chemistry. <https://doi.org/10.1039/c9an01163c>
- [25] Jadvar, H., & Colletti, P. M. (2013). Competitive advantage of PET/MRI [Review of Competitive advantage of PET/MRI]. *European Journal of Radiology*, 83(1), 84. Elsevier BV. <https://doi.org/10.1016/j.ejrad.2013.05.028>
- [26] Kelderhouse, L. E., Chelvam, V., Wayua, C., Mahalingam, S. M., Poh, S., Kularatne, S. A., & Low, P. S. (2013). Development of Tumor-Targeted Near Infrared Probes for Fluorescence Guided Surgery. *Bioconjugate Chemistry*, 24(6), 1075. <https://doi.org/10.1021/bc400131a>
- [27] Khader, A. A., Braschi-Amirfarzan, M., McIntosh, L. J., Gosangi, B., Wortman, J. R., Wald, C., & Thomas, R. (2022). Importance of tumor subtypes in cancer imaging [Review of Importance of tumor subtypes in cancer imaging]. *European Journal of Radiology Open*, 9, 100433. Elsevier BV. <https://doi.org/10.1016/j.ejro.2022.100433>
- [28] Kho, E., Boer, L. L. de, Post, A. L., Vijver, K. V. de, Jóźwiak, K., Sterenborg, H. J. C. M., & Ruers, T. J. M. (2019). Imaging depth variations in hyperspectral imaging: Development of a method to detect tumor up to the required tumor-free margin width. *Journal of Biophotonics*, 12(11). <https://doi.org/10.1002/jbio.201900086>
- [29] Kleijn, B. J. de, Heldens, G. T. N., Herruer, J. M., Sier, C. F. M., Piazza, C., Bree, R. de, Guntinas-Lichius, O., Kowalski, L. P., Poorten, V. V., Rodrigo, J. P., Zidar, N., Nathan, C. O., Tsang, R. K., Golusiński, P., Shaha, A. R., Ferlito, A., & Takes, R. P. (2023). Intraoperative Imaging Techniques to Improve Surgical Resection Margins of Oropharyngeal Squamous Cell Cancer: A Comprehensive Review of Current Literature [Review of Intraoperative Imaging Techniques to Improve Surgical Resection Margins of Oropharyngeal Squamous Cell Cancer: A Comprehensive Review of Current Literature]. *Cancers*, 15(3), 896. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/cancers15030896>
- [30] Koning, S. G. B. de, Baltussen, E. J. M., Karakullukçu, B., Smit, L. A., Veen, R. L. P. van, Hendriks, B. H. W., Sterenborg, H. J. C. M., & Ruers, T. J. M. (2017). Diffuse reflectance spectroscopy from 400-1600 nm to evaluate tumor resection margins during head and neck surgery (Conference Presentation). <https://doi.org/10.1117/12.2251460>
- [31] Krebs, S., Dacek, M. M., Carter, L. M., Scheinberg, D. A., & Larson, S. M. (2020). CAR Chase: Where Do Engineered Cells Go in Humans? *Frontiers in Oncology*, 10. <https://doi.org/10.3389/fonc.2020.577773>
- [32] Krüger-Stokke, B., Bertilsson, H., Langørgen, S., Sjøbakk, T. A. E., Bathen, T. F., & Selnæs, K. M. (2021). Multiparametric Prostate MRI in Biopsy-Naïve Men: A Prospective Evaluation of Performance and Biopsy Strategies. *Frontiers in Oncology*, 11. <https://doi.org/10.3389/fonc.2021.745657>
- [33] Lauwerends, L. J., Abbasi, H., Schut, T. C. B., Driel, P. B. A. A. van, Hardillo, J. A., Santos, I., Barroso, E. M., Koljenović, S., Vahrmeijer, A. L., Jong, R. J. B. de, Puppels, G. J., & Keereweer, S. (2022). The complementary value of intraoperative fluorescence imaging and Raman spectroscopy for cancer surgery: combining the incompatibles [Review of The complementary value of intraoperative fluorescence imaging and Raman spectroscopy for cancer surgery: combining the incompatibles]. *European Journal of Nuclear Medicine and Molecular Imaging*, 49(7), 2364. Springer Science+Business Media. <https://doi.org/10.1007/s00259-022-05705-z>
- [34] Lee, C. H., Tan, T. W., & Tan, C. H. (2021). Multiparametric MRI in Active Surveillance of Prostate Cancer: An Overview and a Practical Approach [Review of Multiparametric MRI in Active Surveillance of Prostate Cancer: An Overview and a Practical Approach]. *Korean Journal of Radiology*, 22(7), 1087. Korean Society of Radiology. <https://doi.org/10.3348/kjr.2020.1224>
- [35] Liao, Z., Lizio, M. G., Corden, C., Khout, H., Rakha, E. A., & Notingher, I. (2020). Feasibility of integrated high-wavenumber Raman imaging and fingerprint Raman spectroscopy for fast margin assessment in breast cancer surgery. *Journal of Raman Spectroscopy*, 51(10), 1986. <https://doi.org/10.1002/jrs.5937>
- [36] Lue, N., Kang, J. W., Yu, C., Barman, I., Dingari, N. C., Feld, M. S., Dasari, R. R., & Fitzmaurice, M. (2012). Portable Optical Fiber Probe-Based Spectroscopic Scanner for Rapid Cancer Diagnosis: A New Tool for Intraoperative Margin Assessment. *PLoS ONE*, 7(1). <https://doi.org/10.1371/journal.pone.0030887>
- [37] Mahalingam, S. M., Kularatne, S. A., Myers, C., Gagare, P. D., Norshi, M., Liu, X., Singhal, S., & Low, P. S. (2018). Evaluation of Novel Tumor-Targeted Near-Infrared Probe for Fluorescence-Guided Surgery of Cancer. *Journal of Medicinal Chemistry*, 61(21), 9637. <https://doi.org/10.1021/acs.jmedchem.8b01115>
- [38] Mahamongkol, K., Teyateeti, A., Woranisarakul, V., Srinualnad, S., & Hansomwong, T. (2025). MRI-guided biopsy reduces biochemical recurrence in prostate cancer patients undergoing radiation therapy: a single-center study from Thailand. *Scientific Reports*, 15(1). <https://doi.org/10.1038/s41598-025-95750-z>
- [39] Malherbe, C., Crutzen, B., Schrooyen, J., Caruso, G., Lecouvet, F., Detrembleur, C., Schubert, T., & Docquier,

- P. (2020). Assessment of Resection Margins in Bone Tumor Surgery. *Sarcoma*, 2020, 1. <https://doi.org/10.1155/2020/5289547>
- [40] Marko, A. J., Borah, B. M., Sifers, K. E., Missert, J. R., Gupta, A., Pera, P., Isaac-Lam, M. F., & Pandey, R. K. (2020). Targeted Nanoparticles for Fluorescence Imaging of Folate Receptor Positive Tumors. *Biomolecules*, 10(12), 1651. <https://doi.org/10.3390/biom10121651>
- [41] Medical imaging in personalised medicine: a white paper of the research committee of the European Society of Radiology (ESR). (2015). *Insights into Imaging*, 6(2), 141. <https://doi.org/10.1007/s13244-015-0394-0>
- [42] Meng, X., Rosenkrantz, A. B., Mendhiratta, N., Fenstermaker, M., Huang, W., Wysock, J., Bjurlin, M. A., Marshall, S., Deng, F.-M., Zhou, M., Melamed, J., Huang, W. C., Lepor, H., & Taneja, S. S. (2015). Relationship Between Prebiopsy Multiparametric Magnetic Resonance Imaging (MRI), Biopsy Indication, and MRI-ultrasound Fusion-targeted Prostate Biopsy Outcomes. *European Urology*, 69(3), 512. <https://doi.org/10.1016/j.eururo.2015.06.005>
- [43] Merriel, S. W. D., Hall, R., Walter, F. M., Hamilton, W., & Spencer, A. (2023). Systematic Review and Narrative Synthesis of Economic Evaluations of Prostate Cancer Diagnostic Pathways Incorporating Prebiopsy Magnetic Resonance Imaging [Review of Systematic Review and Narrative Synthesis of Economic Evaluations of Prostate Cancer Diagnostic Pathways Incorporating Prebiopsy Magnetic Resonance Imaging]. *European Urology Open Science*, 52, 123. Elsevier BV. <https://doi.org/10.1016/j.euros.2023.03.010>
- [44] Munley, M. T., Kagadis, G. C., McGee, K. P., Kirov, A. S., Jang, S., Mutic, S., Jeraj, R., Xing, L., & Bourland, J. D. (2013). An introduction to molecular imaging in radiation oncology: A report by the AAPM Working Group on Molecular Imaging in Radiation Oncology (WGMIR) [Review of An introduction to molecular imaging in radiation oncology: A report by the AAPM Working Group on Molecular Imaging in Radiation Oncology (WGMIR)]. *Medical Physics*, 40(10). Wiley. <https://doi.org/10.1118/1.4819818>
- [45] Nandu, H., Wen, P. Y., & Huang, R. Y. (2018). Imaging in neuro-oncology [Review of Imaging in neuro-oncology]. *Therapeutic Advances in Neurological Disorders*, 11. SAGE Publishing. <https://doi.org/10.1177/1756286418759865>
- [46] Nguyen, T. T., Bhosale, P., Xu, G., Pan, T., Wei, P., & Lü, Y. (2022). Comparison of PSMA-based 18F-DCFPyL PET/CT and pelvic multiparametric MRI for lesion detection in the pelvis in patients with prostate cancer. *PubMed*, 12(6), 166. <https://pubmed.ncbi.nlm.nih.gov/36636233>
- [47] Ong, E. (2018). Preoperative imaging for breast conservation surgery—do we need more than conventional imaging for local disease assessment? [Review of Preoperative imaging for breast conservation surgery—do we need more than conventional imaging for local disease assessment?]. *Gland Surgery*, 7(6), 554. AME Publishing Company. <https://doi.org/10.21037/gs.2018.08.05>
- [48] Ota, Y., Sato, S., Yoshihara, M., Nakamura, Y., Miyagi, E., & Miyagi, Y. (2022). A practical spatial analysis method for elucidating the biological mechanisms of cancers with abdominal dissemination in vivo. *Scientific Reports*, 12(1). <https://doi.org/10.1038/s41598-022-24827-w>
- [49] Perlman, O., Weitz, I. S., & Azhari, H. (2015). Copper oxide nanoparticles as contrast agents for MRI and ultrasound dual-modality imaging. *Physics in Medicine and Biology*, 60(15), 5767. <https://doi.org/10.1088/0031-9155/60/15/5767>
- [50] Pinto, M., Zorn, K. C., Tremblay, J. P., Desroches, J., Dallaire, F., Aubertin, K., Marple, E., Kent, C., Leblond, F., Trudel, D., & Lesage, F. (2019). Integration of a Raman spectroscopy system to a robotic-assisted surgical system for real-time tissue characterization during radical prostatectomy procedures. *Journal of Biomedical Optics*, 24(2), 1. <https://doi.org/10.1117/1.jbo.24.2.025001>
- [51] Predina, J. D., Newton, A. D., Connolly, C., Dunbar, A., Baldassari, M. P., Deshpande, C., Cantu, E., Stadanlick, J., Kularatne, S. A., Low, P. S., & Singhal, S. (2017). Identification of a Folate Receptor-Targeted Near-Infrared Molecular Contrast Agent to Localize Pulmonary Adenocarcinomas. *Molecular Therapy*, 26(2), 390. <https://doi.org/10.1016/j.ymthe.2017.10.016>
- [52] Reichel, D., Curtis, L. T., Ehlman, E., Evers, B. M., Rychahou, P., Frieboes, H. B., & Bae, Y. (2017). Development of Halofluorochromic Polymer Nanoassemblies for the Potential Detection of Liver Metastatic Colorectal Cancer Tumors Using Experimental and Computational Approaches. *Pharmaceutical Research*, 34(11), 2385. <https://doi.org/10.1007/s11095-017-2245-9>
- [53] Roeder, R. K., Curtis, T. E., Nallathamby, P. D., Irimata, L. E., McGinnity, T. L., Cole, L. E., Vargo-Gogola, T., & Dahl, K. D. C. (2017). Nanoparticle imaging probes for molecular imaging with computed tomography and application to cancer imaging. *Proceedings of SPIE, the International Society for Optical Engineering/Proceedings of SPIE*. <https://doi.org/10.1117/12.2255688>

- [54] Salih, S., Elliyaanti, A., Alkatheeri, A., AlYafei, F., Almarri, B., & Khan, H. (2023). The Role of Molecular Imaging in Personalized Medicine [Review of The Role of Molecular Imaging in Personalized Medicine]. *Journal of Personalized Medicine*, 13(2), 369. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/jpm13020369>
- [55] Scimone, M. T., Krishnamurthy, S., Maguluri, G., Preda, D. V., Park, J., Grimbale, J., Song, M. K., Ban, K., & Iftimia, N. (2021). Assessment of breast cancer surgical margins with multimodal optical microscopy: A feasibility clinical study. *PLoS ONE*, 16(2). <https://doi.org/10.1371/journal.pone.0245334>
- [56] Shoman, H., Al-Kassmy, J., Ejaz, M., Matta, J., Alakhra, S., Kahla, K., & D'Acunto, M. (2023). Surgical margin assessment of bone tumours: A systematic review of current and emerging technologies [Review of Surgical margin assessment of bone tumours: A systematic review of current and emerging technologies]. *Journal of Bone Oncology*, 39, 100469. Elsevier BV. <https://doi.org/10.1016/j.jbo.2023.100469>
- [57] Stammes, M. A., Bugby, S. L., Porta, T., Pierzchalski, K., Devling, T., Otto, C., Dijkstra, J., Vahrmeijer, A. L., Geus-Oei, L. de, & Micog, J. S. D. (2018). Modalities for image- and molecular-guided cancer surgery [Review of Modalities for image- and molecular-guided cancer surgery]. *British Journal of Surgery*, 105(2). Oxford University Press. <https://doi.org/10.1002/bjs.10789>
- [58] Szabó, B. K., & Wiberg, M. K. (2006). Dynamic MR imaging as a predictor of prognosis in breast cancer. *Journal of Spectroscopy*, 20(1), 19. <https://doi.org/10.1155/2006/417637>
- [59] Thomas, C. (2011). Functional oncoimaging techniques with potential clinical applications [Review of Functional oncoimaging techniques with potential clinical applications]. *Frontiers in Bioscience-Elite*, 1, 1081. Frontiers Media. <https://doi.org/10.2741/443>
- [60] Thomas, G., Nguyen, T., Pence, I. J., Caldwell, B., O'Connor, M., Giltane, J. M., Sanders, M. E., Grau, A. M., Meszoely, I. M., Hooks, M., Kelley, M. C., & Mahadevan-Jansen, A. (2017). Evaluating feasibility of an automated 3-dimensional scanner using Raman spectroscopy for intraoperative breast margin assessment. *Scientific Reports*, 7(1). <https://doi.org/10.1038/s41598-017-13237-y>
- [61] Vitaz, T. W. (2015). Techniques to Improve the Extent of Brain Tumor Resection — Awake Speech and Motor Mapping, and Intraoperative MRI. In *InTech eBooks*. <https://doi.org/10.5772/58977>
- [62] Wang, K., Chi, C., Hu, Z., Liu, M., Hui, H., Shang, W., Peng, D., Zhang, S., Ye, J., Liu, H., & Tian, J. (2015). Optical Molecular Imaging Frontiers in Oncology: The Pursuit of Accuracy and Sensitivity. *Engineering*, 1(3), 309. <https://doi.org/10.15302/j-eng-2015082>
- [63] Weissleder, R. (2006). Molecular Imaging in Cancer. *Science*, 312(5777), 1168. <https://doi.org/10.1126/science.1125949>
- [64] Wu, M., & Shu, J. (2018). Multimodal Molecular Imaging: Current Status and Future Directions [Review of Multimodal Molecular Imaging: Current Status and Future Directions]. *Contrast Media & Molecular Imaging*, 2018, 1. Hindawi Publishing Corporation. <https://doi.org/10.1155/2018/1382183>
- [65] Yaroslavsky, A. N., Neel, V., & Anderson, R. R. (2003). Demarcation of Nonmelanoma Skin Cancer Margins in Thick Excisions Using Multispectral Polarized Light Imaging. *Journal of Investigative Dermatology*, 121(2), 259. <https://doi.org/10.1046/j.1523-1747.2003.12372.x>
- [66] You, J., Pan, C., Park, K., Li, A., & Du, C. (2019). In vivo detection of tumor boundary using ultrahigh-resolution optical coherence angiography and fluorescence imaging. *Journal of Biophotonics*, 13(3). <https://doi.org/10.1002/jbio.201960091>
- [67] Young, K., Ma, E., Kejiwal, S., Nielsen, T., Aulakh, S. S., & Birkeland, A. C. (2022). Intraoperative In Vivo Imaging Modalities in Head and Neck Cancer Surgical Margin Delineation: A Systematic Review [Review of Intraoperative In Vivo Imaging Modalities in Head and Neck Cancer Surgical Margin Delineation: A Systematic Review]. *Cancers*, 14(14), 3416. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/cancers14143416>
- [68] Yu, H., & Zhang, X. (2020). Synthesis of Prostate MR Images for Classification Using Capsule Network-Based GAN Model. *Sensors*, 20(20), 5736. <https://doi.org/10.3390/s20205736>
- [69] Zheng, D., He, X., & Jing, J. (2023). Overview of Artificial Intelligence in Breast Cancer Medical Imaging [Review of Overview of Artificial Intelligence in Breast Cancer Medical Imaging]. *Journal of Clinical Medicine*, 12(2), 419. Multidisciplinary Digital Publishing Institute. <https://doi.org/10.3390/jcm12020419>
- [70] Zhou, Y., Liu, C., Li, J., Li, Z., Zhou, L., Chen, K., Pu, Y., He, Y., Zhu, K., Li, Q., & Alfano, R. R. (2014). Tumor margin detection using optical biopsy techniques. *Proceedings of SPIE, the International Society for Optical Engineering/Proceedings of SPIE*, 8940, 894014. <https://doi.org/10.1117/12.2038723>