

Pharmacological Assessment of Morinda Citrifolia Fruit Extract's in Anti-Arthritic Properties in Wistar Rats

Rajni Yadav¹, Saumya Malaiya², Dr Shweta Mishra³, Parveen Nisha⁴, Aarekh Kumar Jain⁵, Anshul Chaubey⁶, Basant Khare^{7*}

¹Amity Institute of Pharmacy, Amity University, Manth (Kharora), State Highway 9, Raipur, CG, 493225

²Assistant professor, Adina Institute of Pharmaceutical Science, NH, Bhopal Road, Sagar (M.P.), 470001

³Professor & Head of department Guru Ramdas Khalsa institute of science and technology (Pharmacy), Barela, Jabalpur, MP, 483001

⁴Associate Professor, Vedic Institute of Pharmaceutical Education and Research, Link Road, Bahupura, Sagar, MP, 470003

⁵School of Pharmaceutical Sciences, RGPV Campus, Gandhi Nagar, Bhopal, MP, 462033

⁶Shanti College Of Pharmacy, Chobara, Nowgong, Mudwara Road, Chhatarpur, MP, 471201

⁷Adina College of Pharmacy, ADINA Campus Rd, Lahdara, Sagar, MP, 47000

*Corresponding Author:

Associate professor

Adina College of Pharmacy, ADINA Campus Rd, Lahdara, Sagar, MP, 470001,

Email ID: basant.khare08@gmail.com

Cite this paper as: Rajni Yadav, Saumya Malaiya, Dr Shweta Mishra, Parveen Nisha, Aarekh Kumar Jain, Anshul Chaubey, Basant Khare, (2025) Pharmacological Assessment of Morinda Citrifolia Fruit Extract's in Anti-Arthritic Properties in Wistar Rats. *Journal of Neonatal Surgery*, 14 (4s), 181-189.

ABSTRACT

Arthritis and related disorders, including rheumatoid arthritis (RA), are common diseases affecting millions of people. RA is characterized by articular injuries having an inflammatory propagation of synovial cells, attaining a nearly complete functional defect. It affects about 1% of the general population. *Morinda citrifolia* L (Rubiaceae), commonly called Noni or Indian mulberry, is a small evergreen tree or shrub of Polynesian origin. *M. citrifolia* bears a lumpy, green to yellowish-white fruit, normally 5 to 10 cm in length, with a surface covered in polygonal-shaped sections. The leaves, roots, bark and fruits have all been used medicinally to treat a wide range of ailments. These include, but are not limited to, diabetes, diarrhoea, hypertension, malaria, pain, and topical infections. The fruits are also eaten as a food, but primarily only in times of famine. The present study was designed to investigate anti-arthritic activity of methanolic fruits extract of *M. citrifolia* in Freund's complete adjuvant (FCA)-induced arthritis in rats. Qualitative analysis of various phytochemical constituents was determined by the well-known test protocol available in the literature. The *In vitro* antioxidant activity of methanolic extract of the fruits was assessed against DPPH free radical scavenging assay method using standard protocols. *M. citrifolia* was evaluated for anti-arthritic action by Freund's adjuvant induced arthritis test in adult Albino rats (200±20gm). Rats were injected 0.1 ml of complete Freund's adjuvant into the planter region of the left hind paw. *M. citrifolia* extracts (200 and 400 mg/kg, b.wt.) was given orally to arthritic rats induced with Complete Freund's Adjuvant and changes in rat paw volume and body weight was determined. Phytochemical analysis of methanolic extract of *M. citrifolia* showed the presence of carbohydrate, alkaloids, phenolics, tannin, triterpenoids, steroids, flavonoids and glycoside. In the oral acute toxicity studies, *M. citrifolia* was found to be safe as it did not cause any mortality up to 2000 mg/kg. Hence, 200 and 400 mg/kg doses were selected for the present study. DPPH radical scavenging activity of *M. citrifolia* extract exhibited percent inhibition 60.15% and its IC₅₀ value was found to be 57.10 µg/ml. Ascorbic was used as a reference compound which exhibited percent inhibition 86.36% and showed IC₅₀ value of 19.36 µg/ml. *M. citrifolia* administered groups showed marked reduction in paw volume when compared with the negative control group (Group II). It was also found that there was significant weight loss when compared to standard. The results reveal promising anti-arthritic potential of the *M. citrifolia*. However further pharmacological investigation using isolated active ingredients can be carried out to confirm its efficacy and mechanism of action.

Keywords: *Morinda citrifolia*, Anti-arthritic activity, Freund's adjuvant induced, Phytochemical analysis, *In vitro* antioxidant activity

1. INTRODUCTION

Rheumatoid arthritis (RA) is a systemic autoimmune disease of unknown etiology, affecting 1-1.5% of the population worldwide [1]. The disease is characterized by articular inflammation and by the formation of an inflammatory and invasive tissue, rheumatoid pannus that eventually leads to the destruction of joints. Anti-inflammatory drugs and analgesics including steroids are used to suppress the symptoms, while disease-modifying antirheumatic drugs (DMARDs), newer therapies such as anti-CD20 therapy (rituximab) and abatacept, anti-tumor necrosis factor (TNF)- α therapy (etanercept, infliximab and adalimumab) are often required to inhibit or halt the underlying immune process. However, all of these agents are associated with numerous side effects. In recent days, researchers are directed toward traditional system of medicine for the discovery of drugs that are long-acting anti-inflammatory with minimum side effects. Although there is no ideal animal model for RA at this time, rat adjuvant arthritis shares many features of human RA and the sensitivity of this model to anti-arthritic agents support the view that the adjuvant arthritis is the best available model of RA [2]. Herbal medicines are being accepted and used increasingly by general populations in both eastern and western countries because of the ethnic acceptability and compatibility having fewer side effects [3]. *M. citrifolia* (Rubiaceae), popularly known as Indian mulberry or noni, is a plant indigenous to India, Burma, China, and the Polynesian islands [4, 5]. *M. citrifolia* fruits have been used traditionally by native Polynesians to treat diabetes, high blood pressure, cancer, injury, arthritis, digestive distress, arteriosclerosis, pain, and senility [6]. In addition, *M. citrifolia* fruits have also been used as a remedy for halitosis, bacterial and helminthic infection, wound healing, menstrual cramps, arthritis, gastric and oral ulcers, toothache, and indigestion. It improves lactation and also acts as a purgative [6]. Several researchers have confirmed that the immunostimulant properties of *M. citrifolia* significantly contribute to its antitumor potential [6]. Recently, Palu et al. (2008) demonstrated that *M. citrifolia* juice concentrate (NFJC) stimulated the cannabinoid receptors (CB2). In addition, a decrease in interleukin-4 (IL-4) levels with a concomitant increase in interferon- γ (IFN- γ) levels elicited by NFJC indicated the immunostimulant properties of this plant [7]. To date, the major chemical constituents of this plant have been found to be anthraquinones, flavonol glycosides, Iridoid glycosides, lipid glycosides and triterpenoids [8]. Hence, the aim of this study is to prove the therapeutic potential of the plant *M. citrifolia* as an anti-arthritic agent against Freund's complete adjuvant (FCA) induced arthritis.

2. MATERIALS AND METHODS

Collection of plant material

The medicinal plant *M. citrifolia* (300 gm) was collected locally from Bhopal, MP. After cleaning, plant parts were dried under shade at room temperature for 3 days and then in oven dried at 45°C till complete dryness. Dried plant parts were stored in air tight glass containers in dry and cool place to avoid contamination and deterioration. *M. citrifolia* was authenticated by a plant taxonomist in order to confirm its identity and purity.

Chemicals and reagents

Complete Freund's adjuvant (CFA) was procured from Sigma Aldrich chemicals Pvt. Ltd, Hyderabad, India. Indomethacin was obtained from Akums Drugs and Pharmaceuticals, India. All other chemicals used in this study were obtained from Hi Media Laboratories Pvt. Ltd. (Mumbai, India), SRL Pvt. Ltd. (Mumbai, India) and Merck Life Sci. Private Ltd. (Mumbai, India). All other chemicals used in this study were obtained commercially and were of analytical grade and triple distilled water was used for whole experiment was generated in house.

Extraction of plant material

Hot soxhlet extraction method

This technique involved gathering, correctly washing, and properly rinsing the blossoms of *M. citrifolia*. They were mechanically pulverised after being shade-dried. The plant material from *M. citrifolia*, either whole or coarsely powdered, was successively extracted using solvents such as petroleum ether and methanol for various lengths of time. The Soxhlet apparatus' chamber was filled with powder using a "thimble" design. The solvent used for extraction was heated in flasks, and its vapours were then condensed in a condenser. The powder is extracted by touch when the condensed extractant is dropped into the thimble holding it. The liquid inside the chamber syphon drops into the flask when the liquid level in the chamber reaches the top of the syphon tube. This procedure was continued until an evaporated drop of solvent from the syphon tube did not leave any residue. The resulting extract was filtered, dried by concentration, weighed, and stored for later use [9]. The following formula is used to determine the extract's yield.

Yield (%) = Weight of the residue obtained \times 100

Weight of the plant material taken

Phytochemical screening of the extract

A variety of phytoconstituents, including alkaloids, carbohydrates, glycosides, phytosterols, saponins, tannins, proteins, amino acids, and flavonoids were qualitatively analysed in the fruits extracts of *M. citrifolia* [10, 11].

In vitro anti oxidant activity of plant extract

DPPH assay

Free radical scavenging activity of the extracts of *M. citrifolia* fruits, based on the scavenging activity of the stable free radical 1,1-diphenyl-2-picrylhydrazyl (DPPH) was determined by the method of Ali et al [12]. Different volume of extracts/standard (20-100 µg/ml) was taken from stock solution in a set of test tubes and methanol was added to make the volume to 1 ml. To this, 2 ml of 0.1 mM DPPH reagent was added and mixed thoroughly. Absorbance at 517 nm was determined after 30 min and the percentage inhibition activity was calculated by using the equation: % scavenging activity = $[(A_0 - A_1)/A_0] \times 100$. Where A₀ is the absorbance of the control and A₁ is the absorbance of the extract. Lower the absorbance, the higher is the free radical scavenging activity. The curves were prepared and the IC₅₀ value was calculated using linear regression analysis.

Animals

Animals will select randomly from animal house of Pinnacle Biomedical Research Institute (PBRI), Bhopal, India and further divided into various treatment groups randomly and kept in propylene cage with sterile husk as bedding. Relative humidity of 30.7 % at 22±2°C and 12:12 light and dark cycle will be maintained in the animal house and fed with standard pellets (Golden Feeds, New Delhi, India) and water will available *ad libitum*. Rats will acclimatize to laboratory conditions for 7 days before carrying out the experiments. Separate group (n=6) of rats will used for each set of experiments. Animal experiments will approve by Institutional Animal Ethics Committee (IAEC) of Pinnacle Biomedical Research Institute (PBRI) Bhopal.

Animals used

- Strain - Albino Wistar rats
- Age - 5-6 weeks
- Sex - either sex
- Body weight - 200±20 gm

Acute oral toxicity

Acute toxicity study of the prepared seed extracts of *M. citrifolia* was carried out according to the Organization for Economic Co-Operation and Development (OECD) Guidelines-423 [13] the animals were fasted for 4 h, but allowed free access to water throughout. As per the OECD recommendations, the starting dose level should be that which is most likely to produce mortality in some of the dosed animals; and when there is no information available on a substance to be tested in this regard; for animal welfare reasons, The dose level to be used as the starting dose is selected from one of four fixed levels 5, 50, 300 and 2000 mg/kg body weight. Acute toxicity was determined as per reported method [14].

Induction of arthritis

Arthritis was induced in rats by the intraplantar injection of 0.1 ml of Complete Freund's Adjuvant (CFA) in the left hind paw. The adjuvant contained heat killed Mycobacterium tuberculosis in sterile paraffin oil (10 mg/ml). The paw volume of all the animal groups was measured at 0, 4, 8 and 12 days after the injection of Freund's complete adjuvant [15].

Freund's adjuvant induced arthritic model

Freud's adjuvant induced arthritis model was used to assess the anti-arthritic activity in albino rats. Animals were divided into five groups of six animals each. Group I served as normal control, Group II served as arthritic control, injected with 0.01 ml Freud's adjuvant i.p., Group III served as reference standard, which received 10 mg/kg body weight IP of indomethacin, Group IV received the extract of *M. citrifolia* the dose of 200mg/kg body weight p.o. and Group V received the extract of *M. citrifolia* the dose of 400mg/kg body weight p.o., respectively.

Design of experiment

| S.No. | Groups | Number of animals | Dose |
|-------|--------------------------------------|-------------------|-----------|
| 1 | Normal control | 6 | 0.2 ml |
| 2 | Arthritic control (Freud's adjuvant) | 6 | 0.1 ml |
| 3 | Reference standard (Indomethacin) | 6 | 10 mg/kg |
| 4 | <i>M. citrifolia</i> extract | 6 | 200 mg/kg |
| 5 | <i>M. citrifolia</i> extract | 6 | 400 mg/kg |

Evaluation of arthritis

Bodyweight examination

During treatment, the change in body weight was measured with the help of digital weighing balance from the day of CFA immunization and then subsequently on 0th, 4th, 8th, and 12st days [16].

Paw thickness (Joint diameter)

Joint diameter was measured using a digital vernier caliper (Mitutoyo digimatic caliper, Japan) before adjuvant administration. The joint diameter was measured again on day (0, 4, 8, and 12).

Statistical analysis

The data is expressed as mean \pm Standard Deviation (SD). Results were analyzed using one-way ANOVA followed by Bonferroni test. Differences were considered as statistically significant at $P < 0.05$, $P < 0.001$ when compared with control.

3. RESULTS

The percentage yields of methanolic extract obtained from *M. citrifolia* are depicted in the table 1. Preliminary phytochemical studies of the extract were done according to the published standard methods. These tests were broad in scope and used to determine the presence of carbohydrate, alkaloids, phenolics, tannin, triterpenoids, steroids, flavonoids and glycoside Table 2. In the present investigation, the *in vitro* anti-oxidant activity of extracts of *M. citrifolia* was evaluated by DPPH radical scavenging activity. The results are summarized in Table 3. To determine the safety of *M. citrifolia* for human use, toxicological evaluation is carried out in experimental animals. In the acute toxicity study, no signs of toxicity were found up to the dose of 2000 mg/kg body weight. Hence 1/10th and 1/5th doses i.e. 200 mg/kg and 400 mg/kg have been fixed as ED₅₀ for present study Table 4. Paw swelling is an index of measuring the anti-arthritis activity of *M. citrifolia* at the dose level 200 & 400 mg/kg, p.o. *M. citrifolia* administered groups showed marked reduction in paw volume when compared with the Negative control group (Group II). It was also found that there was significant weight loss when compared to standard Table 5, 6 & Figure 1, 2.

Table 1: Percentage yield of crude extracts of *M. citrifolia* extract

| S. No | Plant name | Solvent | Theoretical weight | Yield(gm) | % yield |
|-------|----------------------|------------|--------------------|-----------|---------|
| 1 | <i>M. citrifolia</i> | Pet. ether | 300 | 1.21 | 0.40% |
| 2 | | Methanol | 283.02 | 5.99 | 2.11% |

Preliminary phytochemical study

Table 2: Phytochemical testing of extract

| S. No. | Experiment | Presence or absence of phytochemical test | |
|--------|-----------------------|---|--------------------|
| | | Pet. ether extract | Methanolic extract |
| 1. | Alkaloids | | |
| 1.1 | Dragendroff's test | Absent | Present |
| 1.2 | Mayer's reagent test | Absent | Present |
| 1.3 | Wagner's reagent test | Absent | Present |
| 1.3 | Hager's reagent test | Absent | Present |
| 2. | Glycoside | | |
| 2.1 | Borntrager test | Present | Present |
| 2.2 | Legal's test | Present | Present |
| 2.3 | Killer-Killiani test | Present | Present |
| 3. | Carbohydrates | | |

| | | | |
|-----|--|---------|---------|
| 3.1 | Molish's test | Absent | Present |
| 3.2 | Fehling's test | Absent | Present |
| 3.3 | Benedict's test | Absent | Present |
| 3.4 | Barfoed's test | Absent | Present |
| 4. | Proteins and Amino Acids | | |
| 4.1 | Biuret test | Present | Absent |
| 5. | Flavonoids | | |
| 5.1 | Alkaline reagent test | Absent | Present |
| 5.2 | Lead Acetate test | Absent | Present |
| 6. | Tannin and Phenolic compounds | | |
| 6.1 | Ferric Chloride test | Absent | Present |
| 7. | Saponins | | |
| 7.1 | Foam test | Present | Absent |
| 8. | Test for Triterpenoids and Steroids | | |
| 8.1 | Salkowski's test | Present | Present |
| 8.2 | Libbermann-Burchard's test | Present | Present |

***In vitro* anti oxidant activity**

Table 3: DPPH assay of ascorbic acid and methanolic extract

| S. No. | Conc. (µg/ml) | Ascorbic acid (% Inhibition) | methanolic Extract (% Inhibition) |
|-------------|---------------|------------------------------|-----------------------------------|
| 1. | 20 | 52.757 | 42.919 |
| 2. | 40 | 56.919 | 46.871 |
| 3. | 60 | 65.660 | 49.835 |
| 4. | 80 | 71.279 | 53.238 |
| 5. | 100 | 86.368 | 60.153 |
| IC 50 Value | | 19.36 | 57.10 |

Table 4: Acute oral toxicity of *M. citrifolia* extract

| S. No. | Groups | Observations/ Mortality |
|--------|-----------------------|-------------------------|
| 1. | 5 mg/kg Bodyweight | 0/3 |
| 2. | 50 mg/kg Bodyweight | 0/3 |
| 3. | 300 mg/kg Bodyweight | 0/3 |
| 4. | 2000 mg/kg Bodyweight | 0/3 |

Freund's adjuvant induced arthritic model


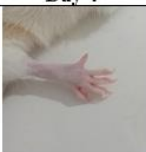







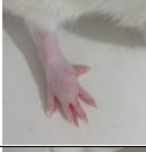



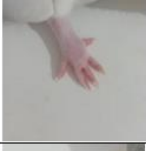


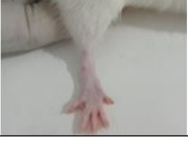


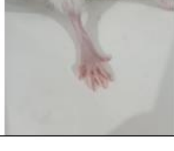
Table 5: Body weight

| Body weight | | |
|---|----------------|--------------|
| | Initial weight | Final weight |
| Normal control | 210±1.62 | 216±2.89 |
| Arthritic control (Freud's adjuvant) | 211±1.66 | 221±2.49 |
| standard (Indomethacin) 10 mg/kg | 201±0.18 | 213±1.30 |
| <i>M. citrifolia</i> extract(200 mg/kg) | 206±1.05 | 215±0.35 |
| <i>M. citrifolia</i> extract(400 mg/kg) | 205±1.52 | 218±7.39 |

Table 6: Freund's adjuvant induced arthritic Model

| Rat paw volume | | | | |
|---|-----------|-----------|-----------|--------------|
| Groups | Day 0 | Day 4 | Day 8 | Day 12 |
| Normal control | 0.23±0.05 | 0.25±0.10 | 0.26±0.09 | 0.25.99±0.08 |
| Arthritic control (Freud's adjuvant) | 0.28±0.07 | 0.71±0.08 | 0.77±0.12 | 0.80±0.09 |
| Standard (Indomethacin) 10 mg/kg | 0.25±0.01 | 0.53±0.03 | 0.42±0.07 | 0.40±0.21 |
| <i>M. citrifolia</i> extract(200 mg/kg) | 0.27±0.15 | 0.59±0.21 | 0.48±0.13 | 0.46±0.24 |
| <i>M. citrifolia</i> extract(400 mg/kg) | 0.26±0.04 | 0.54±0.11 | 0.45±0.19 | 0.44±0.17 |

Table 7: Histology of anti arthritic activity

| Group | Day 0 | Day 4 | Day 8 | Day 12 |
|---|---|---|--|---|
| I. Normal control |  |  |  |  |
| II. Arthritic control (freud's adjuvant) |  |  |  |  |
| III. standard (Indomethacin) 10 mg/kg |  |  |  |  |
| IV. <i>Morinda citrifolia</i> extract(200 mg/kg) |  |  |  |  |
| V. <i>Morinda citrifolia</i> extract(400 mg/kg) |  |  |  |  |

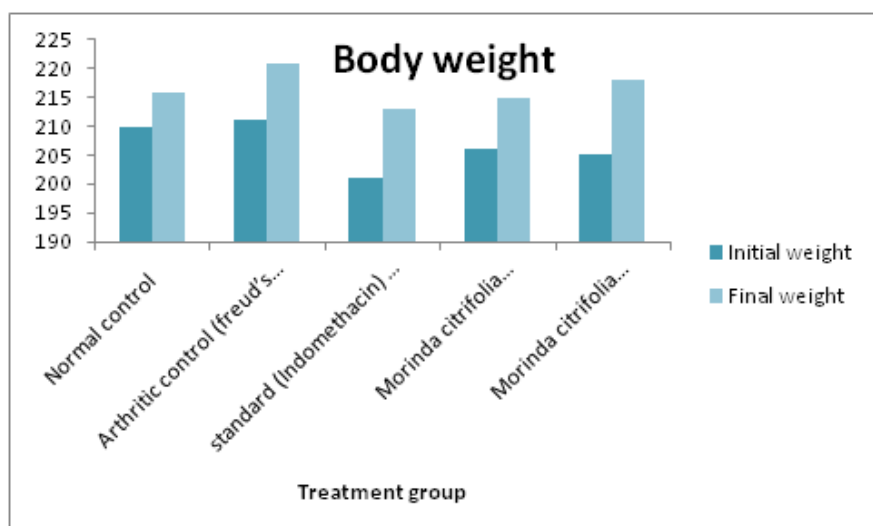


Figure1: Represent the body weight

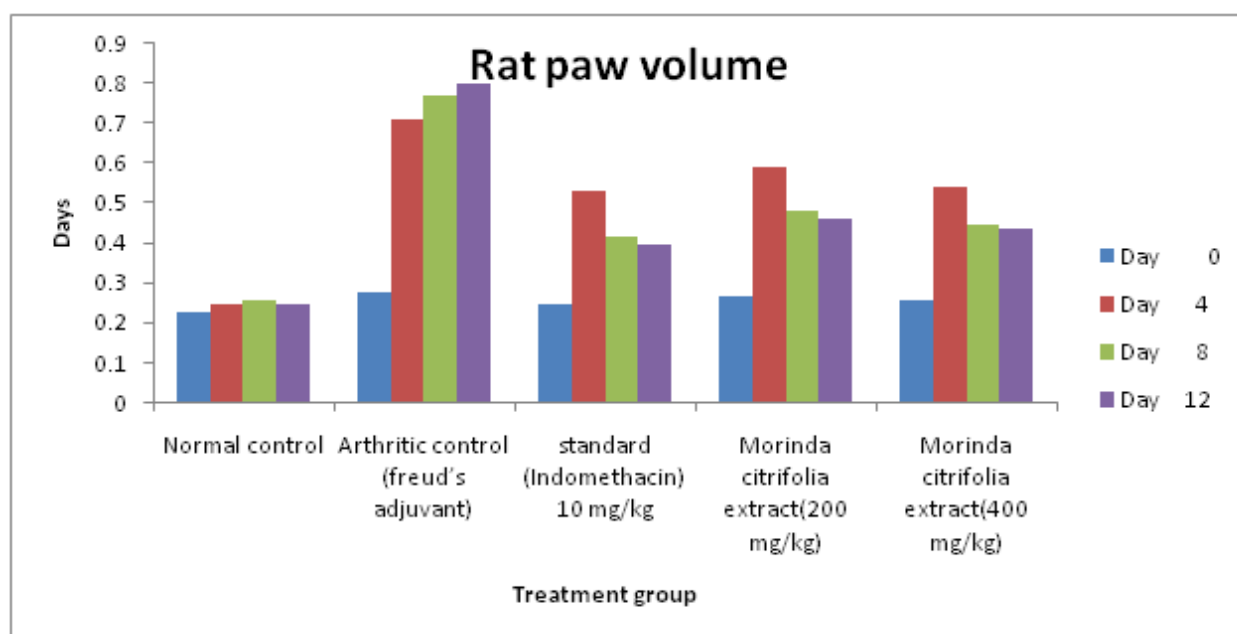


Figure2: Rat paw volume

4. DISCUSSION

Rheumatoid arthritis is an autoimmune disorder, the immunologically mediated complete Freund's adjuvant induced arthritic model of chronic inflammation is considered as the best available experimental model of rheumatoid arthritis. Complete Freund's adjuvant-induced arthritis is a model of chronic polyarthritis with features that resemble rheumatoid arthritis [17]. The determination of paw swelling is apparently simple, sensitive and quick procedure for evaluating the degree of inflammation and assessing of therapeutic effects of drugs [18]. In adjuvant-induced arthritis model rats developed a chronic swelling in multiple joints with influence of inflammatory cells, erosion of joint cartilage and bone destruction and remodeling which have close similarities to human rheumatoid disease. These inflammatory changes ultimately result in the complete destruction of joint integrity and functions in the affected animal. Also, the CFA administered rats showed soft tissue swelling around the ankle joints during the development of arthritis, which was considered as edema of the particular tissues [19].

In our study the methanolic extract of *M. citrifolia* exhibited a significant anti-arthritic activity in a dose dependent manner. In the present study, we showed that methanolic extract of *M. citrifolia* could significantly inhibit the progression of the rheumatoid arthritis in treated animals. However, standard drug and methanolic extract significantly suppressed the swelling of the paws in both acute and chronic phase which may be due to the suppression of inflammatory mediator released due to

induction of Freund's adjuvant. Though the actual mechanism of suppressing inflammation is not known but it can be correlated with the presence of flavonoids in suppressing the inflammation and anti-oxidant activity. Numerous studies have suggested a role of oxidative stress in the pathogenesis of rheumatoid arthritis [20]. Therefore; it was assumed that the reported and well-established antioxidant properties of *M. citrifolia* and its ability to block the COX-2 pathway during the progression of inflammation justify the usage of the plant extract in the treatment of rheumatoid arthritis. There was a significant reduction in the paw volume in Freund's complete adjuvant induced arthritic rats. The cardinal signs of the chronic inflammatory reactions like redness, swelling, arthralgia and immobility of affected joints were significantly less in the drug treated animal than those of the control. The pathogenesis or reasons for development of arthritis following injection of FCA are not fully understood. The result of the present study also indicates that there is a close relationship between the extent of inflammation, loss of body weight and arthritic index. The arthritic scoring was done on the basis of visual observation where it can be seen that there is a marked reduction in the swelling and joint damage of the drug treated groups [21]. It was also noted that the high dose *Morinda citrifolia*. Extract proved its efficacy to reduce the inflammation of the paws.

5. CONCLUSION

Preliminary phytochemical investigations on the methanolic extract of *M. citrifolia* were noted the presence of carbohydrates, flavonoids, saponins, alkaloids and glycosides. No mortality or behavioral abnormality recorded in mice during experiments at the highest dose level of 2000 mg/kg tested for LD50 studies. The high dose of methanolic extract of *M. citrifolia* exhibited a significant anti-arthritis activity by reducing Paw volumes and increase in body weight. Phytochemical constituents like flavonoids, saponins, glycosides and alkaloids were already reported for their anti-arthritis activity and these constituents were present in methanolic extract of *M. citrifolia*. Hence these chemical constituents can be accounted for the observed anti-arthritis activities.

REFERENCES

- [1] Liu M, Dong J, Yang Y, Yang X, Xu H. Anti-inflammatory effects of triptolide loaded poly (D, L-lactic acid) nanoparticles on adjuvant induced arthritis in rats. *J Ethnopharmacol* 2005; 97:219-25.
- [2] Ward JR, Cloud RS. Comparative effect of antirheumatic drugs on adjuvant induced polyarthritis in rats. *J Pharmacol Exp Ther* 1966; 152:116-21.
- [3] Balasubramanian A, Manivannan R, Paramaguru R, Mazudmer PM, Vijayakumar M. Evaluation of anti-inflammatory and antioxidant activities of stem bark of *Toddalia asiatica* (L.) Lam using different experimental models. *Pharmacologia* 2012; 3:144-9.
- [4] Kirtikar K, Basu B (1933): *Indian Medicinal Plants*, Vol. II. Delhi, Periodical Expert Book Agency, pp. 1295.
- [5] Wang M, West B, Jensen C, Nowicki D, Su C, Palu A, Anderson G (2002): *Morinda citrifolia* (noni): A literature review and recent advances in noni research. *Acta Pharmacol Sin* 23: 1127–1141.
- [6] Nayak S, Mengi S. Immunostimulant activity of noni (*Morinda citrifolia*) on T and B lymphocytes. *Pharmaceutical biology*. 2010; 48(7):724-31.
- [7] Palu A, Hirazumi A, West B, Deng S, Jensen J, White L (2008): The effects of *Morinda citrifolia* L. (noni) on the immune system: Its molecular mechanisms of action. *J Ethnopharmacol* 115: 502–506.
- [8] Su BN, Pawlus AD, Jung HA, Keller WJ, McLaughlin JL, Kinghorn AD. Chemical Constituents of the Fruits of *Morinda citrifolia* (Noni) and Their Antioxidant Activity. *Journal of natural products*. 2005; 68(4):592-5.
- [9] Pradhan A, Jain P, Pal M, Chauhan M, Jain DK. Qualitative and quantitative determination of phytochemical contents of hydroalcoholic extract of *Salmaliamalabarica*. *Pharmacologyonline* 2019; 1:21-6.
- [10] Dutta R, Sharma MK, Khan A, Jha M. Phytochemical and in vitro antioxidant assay of *Fumaria officinalis* leaf extract. *Journal of Advanced Scientific Research* 2020; 11(03):176-82.
- [11] Jain DK, Gupta S, Jain R, Jain N. Anti-inflammatory Activity of 80% Ethanolic Extract of *Acorus calamus* Linn. Leaves in Albino Rats. *Research Journal of Pharma Technology* 2010; 3(3): 882-884.
- [12] Ali, M. S., Amin, M. R., Kamal, C. M. I., & Hossain, M. A.. In vitro antioxidant, cytotoxic, thrombolytic activities and phytochemical evaluation of methanol extract of the *A. philippense* L. leaves. *Asian Pacific Journal of Tropical Biomedicine* 2013. 3(6), 464-469.
- [13] Guideline Document on Acute oral Toxicity Testing, Series on Testing and Assessment No. 423. Paris: Organization for Economic Co-Operation and Development, OECD Environment, Health and Safety Publications; 1996. Available from: <http://www.oecd.org/ehs>.
- [14] Jonsson M, Jestoi M, Nathanail AV, Kokkonen UM, Anttila M, Koivisto P, Peltonen K. Application of OECD Guideline 423 in assessing the acute oral toxicity of moniliformin. *Food Chem Toxicol*. 2013; 53: 27-32.
- [15] Newbould BB: Chemotherapy of arthritis induced in rats by injection of Mycobacterial adjuvant. *Br J*

PharmacolChemother. 1963, 21: 127-137.

- [16] Patel P, Patel D, Patel N. Experimental investigation of the anti-rheumatoid activity of *Pleurotus sajorcaju* in adjuvant-induced arthritic rats. Chin J Nat Med. 2012; 10:269e274.
 - [17] Pathak N, Gohil P, Patel NB, Kasture S, Jivani N, Bhalodia Y. Curative effect of *Albizia lebbeck* methanolic extract against adjuvant arthritis – with special reference to bone erosion. Int J Pharma Sci Drug Res 2009;1:183-7.
 - [18] Singh S, Majumdar DK. Effect of fixed oil of *Ocimum sanctum* against experimentally induced arthritis and joint edema in laboratory animals. Int J Pharmacog 1996;34:218-22.
 - [19] Yende SR, Sannapuri VD, Vyawahare NS, Harle UN. Antirheumatoid activity of aqueous extract of *Piper nigrum* on Freund's Adjuvant-induced arthritis in rats. Int J Pharma Sci Res 2010;1:129-33.
 - [20] Bauerová K, Bezek A. Role of reactive oxygen and nitrogen species in etiopathogenesis of rheumatoid arthritis. Gen Physiol Biophys 1999;18:15-20.
 - [21] Gelderman KA, Hultqvist M, Olsson LM, Bauer K, Pizzolla A, et al. (2007) Rheumatoid arthritis: the role of reactive oxygen species in disease development and therapeutic strategies. Antioxid Redox Signal 9: 1541-1567.
-